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# Extraction and DFT study on the complexation of Mg<sup>2+</sup> with valinomycin

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**Abstract** From extraction experiments in the two-phase water–nitrobenzene system and  $\gamma$ -activity measurements, the stability constant of the valinomycin–magnesium complex in nitrobenzene saturated with water was determined. Further, the structure of the resulting complex was indicated by means of the density functional level of theory calculations.

**Keywords** Antibiotics · Macrocycles · Stability constant · Ab initio calculations · Complex structure

# Introduction

Valinomycin (1, Scheme 1) is a dodecadepsipeptide made up of three identical tetradepsipeptides, each of which consists of four sub-units of alternating amino acid and hydroxy acid residues, linked by peptide and ester bonds. The valinomycin molecule thus takes the form of a

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P. Vaňura Department of Analytical Chemistry, Faculty of Chemical Engineering, Institute of Chemical Technology, Prague, Czech Republic 36-membered ring and has a multiplicity of conformational possibilities. The ability of valinomycin to carry ions across a membrane is primarily due to its forming of molecular complexes with them and, secondarily, to the lipophilic nature of the outer rim of its depsipeptide ring, which secures its embedding into the membrane. It was originally believed that a complex was formed selectively with potassium cation, but further studies have shown that other metal ions also bind to valinomycin [1–4].

Experimental evidences for a valinomycin–proton complex and for unusual divalent cation complexes of valinomycin have recently been reported [5, 6]. However, up to now, the structure of the valinomycin–magnesium complex has not been elucidated. In this work, the stability constant of the  $1 \cdot Mg^{2+}$  complex in the organic phase of the water–nitrobenzene extraction system was evaluated. Moreover, by applying quantum mechanical density functional level of theory (DFT) calculations, the most probable structure of the above-mentioned cationic complex species was predicted.

### **Results and discussion**

#### Extraction experiments

Results from previous papers [7–11] indicate that the twophase water–Mg(NO<sub>3</sub>)<sub>2</sub>/nitrobenzene–Sr(DCC)<sub>2</sub>–1 (valinomycin) extraction system, chosen for determination of the stability constant of the complex  $1 \cdot Mg^{2+}$  in nitrobenzene saturated with water, can be characterized by the main chemical equilibrium given by Eq. (1), to which the equilibrium extraction constant (Eq. (2)) corresponds; "aq" and "nb" denote the presence of the species in the aqueous and nitrobenzene phases, respectively.



Scheme 1

$$Mg^{2+}(aq) + \mathbf{1} \cdot Sr^{2+}(nb) \rightleftharpoons \mathbf{1} \cdot Mg^{2+}(nb) + Sr^{2+}(aq); K_{ex}(Mg^{2+}, \mathbf{1} \cdot Sr^{2+})$$
(1)

$$K_{\rm ex}(\mathrm{Mg}^{2+},\mathbf{1}\cdot\mathrm{Sr}^{2+}) = \frac{\left[\mathbf{1}\cdot\mathrm{Mg}^{2+}\right]_{\rm nb}\left[\mathrm{Sr}^{2+}\right]_{\rm aq}}{\left[\mathrm{Mg}^{2+}\right]_{\rm aq}\left[\mathbf{1}\cdot\mathrm{Sr}^{2+}\right]_{\rm nb}}$$
(2)

It is necessary to emphasize that **1** is an extremely hydrophobic ligand, practically present in the nitrobenzene phase only, where this ligand forms—with  $Mg^{2+}$  and  $Sr^{2+}$ —the relatively stable complexes  $1 \cdot Mg^{2+}$  and  $1 \cdot Sr^{2+}$ .

Taking into account the conditions of electroneutrality in the organic and aqueous phases of the system under study, the mass balances of Mg<sup>2+</sup> and Sr<sup>2+</sup> ions for equal volumes of the nitrobenzene and aqueous phases, and the measured equilibrium distribution ratio of strontium,  $D_{\rm Sr} = [\mathbf{1}\cdot\mathrm{Sr}^{2+}]_{\rm nb}/[\mathrm{Sr}^{2+}]_{\rm aq}$ , combined with Eq.(2), we obtain the final expression for the above-mentioned extraction constant (Eq. (3));  $C_{\rm Mg(NO_3)_2}^{\rm in,aq}$  is the initial concentration of Mg(NO<sub>3</sub>)<sub>2</sub> in the aqueous phase and  $C_{\rm Sr(DCC)_2}^{\rm in,nb}$ denotes the initial concentration of Sr(DCC)<sub>2</sub> in the organic phase of the system under consideration.

$$K_{\rm ex}({\rm Mg}^{2+}, \mathbf{1} \cdot {\rm Sr}^{2+}) = \frac{1}{D_{\rm Sr}} \frac{C_{\rm Sr(DCC)_2}^{\rm in,nb}}{(1+D_{\rm Sr})C_{\rm Mg(NO_3)_2}^{\rm in,aq} - C_{\rm Sr(DCC)_2}^{\rm in,nb}}$$
(3)

From the extraction experiments and  $\gamma$ -activity measurements, and by using Eq. (3), the value of the constant  $K_{ex}$  (Mg<sup>2+</sup>, **1**·Sr<sup>2+</sup>) was evaluated as log  $K_{ex}$ (Mg<sup>2+</sup>, **1**·Sr<sup>2+</sup>) = -0.4 ± 0.1. Furthermore, in accordance with other results [5, 6, 8], for the exchange extraction constant  $K_{ex}$  (Mg<sup>2+</sup>, Sr<sup>2+</sup>) corresponding to the equilibrium Mg<sup>2+</sup>(aq) + Sr<sup>2+</sup>(nb)  $\rightleftharpoons$  Mg<sup>2+</sup>(nb) + Sr<sup>2+</sup>(aq) and for the extraction constant  $K_{ex}$  (Mg<sup>2+</sup>, **1**·Sr<sup>2+</sup>) defined above, and for the stability constants of the complexes **1**·Sr<sup>2+</sup> and  $1 \cdot Mg^{2+}$  in nitrobenzene saturated with water, denoted by  $\beta_{nb}$  ( $1 \cdot Sr^{2+}$ ) and  $\beta_{nb}$  ( $1 \cdot Mg^{2+}$ ), one obtains Eq. (4).

$$\log \beta_{\rm nb} \left( \mathbf{1} \cdot \mathbf{M} g^{2+} \right) = \log \beta_{\rm nb} \left( \mathbf{1} \cdot \mathbf{S} r^{2+} \right) + \log K_{\rm ex} \left( \mathbf{M} g^{2+}, \mathbf{1} \cdot \mathbf{S} r^{2+} \right) - \log K_{\rm ex} \left( \mathbf{M} g^{2+}, \mathbf{S} r^{2+} \right)$$
(4)

Using the value log  $K_{ex}$  (Mg<sup>2+</sup>, Sr<sup>2+</sup>) = -0.6 inferred from Refs. [10, 11], the constant log  $K_{ex}$  (Mg<sup>2+</sup>, 1·Sr<sup>2+</sup>) given above and log  $\beta_{nb}$  (1·Sr<sup>2+</sup>) = 5.4 [12], and applying Eq. (4), we obtain the stability constant of the 1·Mg<sup>2+</sup> complex species in water-saturated nitrobenzene at 25 °C as log  $\beta_{nb}$ (1·Mg<sup>2+</sup>) = 5.6 ± 0.1. It means that in this medium the stability constants of the complexes 1·Mg<sup>2+</sup> and 1·Sr<sup>2+</sup> are comparable.

## Quantum mechanical calculations

The quantum mechanical calculations were performed at the density functional level of theory (DFT, B3LYP functional) using the Gaussian 03 suite of software [13]. The 6-31G(d) basis set was used and the optimizations were unconstrained. Although a possible effect of the polar solvent on the detailed structures of 1 and the Mg<sup>2+</sup> complex of 1 could be imagined, our quantum mechanical calculations in similar cases, carried out in an analogous way, showed very good agreement of experiment with theory [14–16].

Recently, the hydration number of the valinomycinmagnesium complex in the organic phase of the waternitrobenzene extraction system was determined as  $h(1 \cdot Mg^{2+}) = 2.9 \pm 0.1$  (P. Selucký, 2007, private communication) by means of the method published elsewhere [17]. Hence, in this study, let us consider further both the "nonhydrated" state ( $1 \cdot Mg^{2+}$ ) and the "hydrated" state ( $1 \cdot Mg^{2+} \cdot 3H_2O$ ) of this magnesium complex.

In the model calculations, we optimized the molecular geometry of the parent valinomycin ligand **1** and its complex with  $Mg^{2+}$ . The optimized structure of free **1** having  $C_3$  symmetry, very much like that ingeniously derived by early researchers from their experimental data [18–26], has been presented in our previous paper [27].

In Figs. 1 and 2, two structures ("asymmetrical" and "symmetrical") obtained by full optimizations of the  $1 \cdot Mg^{2+}$  complex are depicted, together with the lengths of the respective  $Mg^{2+}$ ...O bonds (in Å). In this context it is necessary to emphasize that the binding energies corresponding to the "asymmetrical" and "symmetrical" structures of this  $1 \cdot Mg^{2+}$  complex are -1,442.6 and -1,382.3 kJ mol<sup>-1</sup>, respectively. This means that the "asymmetrical" structure of the  $1 \cdot Mg^{2+}$  species shown in Fig. 1 is somewhat energetically favoured (by 62.3 kJ mol<sup>-1</sup>) in comparison with the "symmetrical" structure illustrated in Fig. 2.



Fig. 1 Two projections of the DFT optimized "asymmetrical" structure of the  $1 \cdot Mg^{2+}$  complex [B3LYP/6-31G(d)] (hydrogen atoms are omitted for clarity except for those taking part in internal hydrogen bonds NH···O)

Finally, the optimized structure of the  $1 \cdot Mg^{2+} \cdot 3H_2O$ complex species is presented in Fig. 3. In this complex, the "central" Mg<sup>2+</sup> ion is bound by three strong bonds to three oxygen atoms of the respective water molecules (2.00, 2.02, and 2.05 Å) and to two oxygen atoms of the corresponding C=O groups (2.10 and 2.08 Å) of the parent valinomycin ligand 1. Besides this, the  $1 \cdot Mg^{2+} \cdot 3H_2O$ cationic complex system is evidently stabilized by six strong hydrogen bonds OH…H, as also illustrated in Fig. 3. From this point of view it should be noted that the optimized structure of the 1.Mg<sup>2+</sup>.3H<sub>2</sub>O cationic complex species stabilized by the mentioned hydrogen bonds (Fig. 3) is apparently much more real than the two optimized structures of the complex  $1 \cdot Mg^{2+}$  (Figs. 1, 2). This fact confirms the calculated binding energy of the  $1 \cdot Mg^{2+} \cdot 3H_2O$  complex species (-1,785.7 kJ mol<sup>-1</sup>).



Fig. 2 The DFT optimized "symmetrical" structure of the  $1 \cdot Mg^{2+}$  complex [B3LYP/6-31G(d)] (hydrogen atoms are omitted for clarity except for those taking part in internal hydrogen bonds NH…O)

which is substantially higher than the above-given binding energies corresponding to the "asymmetrical" and "symmetrical" structures of the  $1 \cdot Mg^{2+}$  complex.

## Experimental

Cesium dicarbollylcobaltate (CsDCC) was purchased from Katchem, Řež, Czech Republic. A nitrobenzene solution of HDCC [7] was prepared from CsDCC by the procedure described in Ref. [17]. Equilibration of the nitrobenzene solution of HDCC with a stoichiometric amount of  $Sr(OH)_2$ , which was dissolved in an aqueous solution of  $Sr(NO_3)_2$  (0.2 M), yielded the corresponding  $Sr(DCC)_2$  solution in nitrobenzene. Valinomycin (1) was supplied by Fluka, Buchs, Switzerland. The other chemicals used (Lachema, Brno, Czech Republic) were of reagent-grade purity. The radionuclide  ${}^{85}Sr^{2+}$  (DuPont, Belgium) was of standard radiochemical purity.

The extraction experiments were conducted in 10 cm<sup>3</sup> glass test-tubes with polyethylene stoppers. An aqueous solution of Mg(NO<sub>3</sub>)<sub>2</sub> (2 cm<sup>3</sup>) with a concentration in the range  $1 \times 10^{-3}$  to  $1 \times 10^{-2}$  M and micro amounts of  $^{85}Sr^{2+}$  were added to 2 cm<sup>3</sup> of a nitrobenzene solution of **1** and Sr(DCC)<sub>2</sub>, whose initial concentrations also varied from  $1 \times 10^{-3}$  to  $1 \times 10^{-2}$  M (in all experiments, the initial concentration of **1** in nitrobenzene,  $C_1^{\text{in,nb}}$ , was equal to the initial concentration of Sr(DCC)<sub>2</sub> in this medium,  $C_{\text{Sr(DCC)}_2}^{\text{in,nb}}$ ). The test-tubes filled with these solutions were shaken for 2 h at 25 ± 1 °C, using a laboratory



Fig. 3 Two projections of the DFT optimized structure of the  $1 \cdot Mg^{2+} \cdot 3H_2O$  complex [B3LYP/6-31G(d)] (hydrogen atoms are omitted for clarity except for those of three water molecules and those taking part in internal hydrogen bonds NH…O)

shaker. The phases were then separated by centrifugation. Samples (1 cm<sup>3</sup>) were then taken from each phase and their  $\gamma$ -activities were measured using a well-type NaI(Tl) scintillation detector connected to a NK 350  $\gamma$ -analyzer (Gamma, Budapest, Hungary). The equilibrium distribution ratios of strontium,  $D_{\rm Sr}$ , were determined as the ratios of the measured radioactivities of  ${}^{85}{\rm Sr}^{2+}$  in the nitrobenzene and aqueous samples.

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## References

- 1. Gennis RB (1989) Biomembranes: molecular structure and function. Springer, New York
- 2. Stryer L (1995) Biochemistry. Freeman, New York
- 3. Pressman BC (1976) Ann Rev Biochem 45:501
- 4. Ovchinnikov YA (1979) Eur J Biochem 94:321
- 5. Makrlík E, Vaňura P (2006) Monatsh Chem 137:157
- 6. Makrlík E, Vaňura P, Selucký P (2008) Monatsh Chem 139:597
- 7. Makrlík E, Vaňura P (1985) Talanta 32:423
- Daňková M, Makrlík E, Vaňura P (1997) J Radioanal Nucl Chem 221:251
- 9. Makrlík E, Vaňura P (1998) ACH Models Chem 135:213
- 10. Rais J (1971) Collect Czech Chem Commun 36:3253
- 11. Vaňura P, Makrlík E, Rais J, Kyrš M (1982) Collect Czech Chem Commun 47:1444
- 12. Makrlík E, Vaňura P (2006) J Radioanal Nucl Chem 268:155
- 13. Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Montgomery JA Jr, Vreven T, Kudin KN, Burant JC, Millam JM, Iyengar SS, Tomasi J, Barone V, Mennucci B, Cossi M, Scalmani G, Rega N, Petersson GA, Nakatsuji H, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Klene M, Li X, Knox JE, Hratchian HP, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Ayala PY, Morokuma K, Voth GA, Salvador P, Dannenberg JJ, Zakrzewski VG, Dapprich S, Daniels AD, Strain MC, Farkas O, Malick DK, Rabuck AD, Raghavachari K, Foresman JB, Ortiz JV, Cui Q, Baboul AG, Clifford S, Cioslowski J, Stefanov BB, Liu G, Liashenko A, Piskorz P, Komaromi I, Martin RL, Fox DJ, Keith T, Al-Laham MA, Peng CY, Nanayakkara A, Challacombe M, Gill PMW, Johnson B, Chen W, Wong MW, Gonzalez C, Pople JA (2004) Gaussian 03, Revision C. 02, Gaussian, Inc., Wallingford
- Kříž J, Dybal J, Makrlík E, Vaňura P, Lang J (2007) Supramol Chem 19:419
- Kříž J, Dybal J, Makrlík E, Vaňura P (2008) Supramol Chem 20:387
- Kříž J, Dybal J, Makrlík E, Budka J, Vaňura P (2008) Supramol Chem 20:487
- 17. Makrlík E (1992) Collect Czech Chem Commun 57:289
- Shemyakin MM, Ovchinnikov YA, Ivanov VT, Antonov VK, Vinogradova EI, Shkrob AM, Malenkov GG, Evstratov AV, Laine IA, Melnik EI, Ryabova ID (1969) J Membr Biol 1:402
- Haynes DH, Kowalsky A, Pressman BC (1969) J Biol Chem 244:502
- Ivanov VT, Laine IA, Abdullaev ND, Senyavina LB, Popov EM, Ovchinnikov YA, Shemyakin MM (1969) Biochem Biophys Res Commun 34:803
- 21. Grell E, Funck T (1973) J Supramol Struct 1:307
- 22. Grell E, Funck T, Sauter H (1973) Eur J Biochem 34:415
- 23. Mayers DF, Urry DW (1972) J Am Chem Soc 94:77
- 24. Urry DW, Kumar NG (1974) Biochemistry 13:1829
- Duax WL, Griffin JF, Langs DA, Smith GD, Grochulski P, Pletnev V, Ivanov V (1996) Biopolymers 40:141
- 26. Wang F, Zhao C, Polavarapu PL (2004) Biopolymers 75:85
- 27. Makrlík E, Dybal J, Vaňura P (2009) Monatsh Chem 140:251